Data Collector – Single Cell MRD

**Refer to: BMMC Preparation for Tapestri Single-Cell MRD User Guide**

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| --- | --- | --- | --- |
| **Chapter** | **Step** | **Question** | **Image/Data** |
| **2 |**  Thaw Cells | **A |**  After first centrifugation step | Cell pellet visible?  o **Yes** o **No**  o **Yes** o **No**  o **Yes** o **No** | [15 mL tube with cell pellet] |
| **B |**  After final washing step | Cell pellet visible?  o **Yes** o **No**  o **Yes** o **No**  o **Yes** o **No** | [1.5 mL tube with cell pellet] |
| **C |**  First quantification | Cell concentration quantifiable?  o **Yes** o **No**  o **Yes** o **No**  o **Yes** o **No** | [Cell suspension on slide, exported from imager] |
| **D |**  Quantification prior to cell staining | Cell concentration between 15,000 and 38,500 cells/µL in at least 130 µL?  o **Yes** o **No**  o **Yes** o **No**  o **Yes** o **No** | [Cell suspension on slide, exported from imager, 5X dilution] |
| **4 |**  Enrich Cells | **A |**  After centrifugation with MACS buffer | Cell pellet visible?  o **Yes** o **No**  o **Yes** o **No**  o **Yes** o **No** | [15 mL tube with cell pellet] |
| **5 |**  Multiplex Cells | **A |**  After first centrifugation step | Cell pellet visible?  o **Yes** o **No**  o **Yes** o **No**  o **Yes** o **No** | [1.5 mL tube with cell pellet] |
| **B |**  First quantification | Cell concentration quantifiable?  o **Yes** o **No**  o **Yes** o **No**  o **Yes** o **No** | [Cell suspension on slide, exported from imager] |
| **C |**  After final centrifugation step | Cell pellet visible?  o **Yes** o **No** | [1.5 mL tube with cell pellet] |
| **D |**  Multiplexed sample quantification | Cell concentration quantifiable?  o **Yes** o **No** | [Cell suspension on slide, exported from imager] |
| **E |**  Final quantification after cell dilution | Cell concentration between 2,800 and 3,200 cells/µL?  o **Yes** o **No** | [Cell suspension on slide, exported from imager] |

**Refer to: Tapestri Single Cell DNA + Protein Sequencing v3 User Guide**

*Ensure that instrument firmware is updated for v3 chemistry*

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| --- | --- | --- | --- |
| **Chapter** | **Step** | **Question** | **Image/Data** |
| **2 |**  Encapsulate Cells | **A |**  After reagent loading | Bubbles in any of the reagents?  o **Yes** o **No** | [Top view of DNA cartridge w/o gasket applied] |
| **B |**  After encapsulation program finished | Emulsions fully intact?  o **Yes** o **No** | [Emulsion-safe tube w/ encapsulated cells] |
| **C |**  After encapsulation program finished | Remaining cell suspension volume [reservoir 2] < 10 µL?  o **Yes** o **No** | [Measure volume] |
| **D |**  When removing oil | Total volume ~100 µL in the tube?  o **Yes** o **No** | [Emulsion-safe tube w/ encapsulated cells and oil removed] |
| **3 |**  Lyse and Digest Cells | **A |**  Before thermal cycling | Thermal cycling protocol correct?  o **Yes** o **No** | [Thermal cycling protocol image] |
| **B |**  Before thermal cycling | PCR skirt used?  o **Yes** o **No** | [Thermal heat block image] |
| **C |**  After thermal cycling | Emulsions fully intact?  o **Yes** o **No** | [Emulsion-safe tube w/ encapsulated cells] |
| **4 |**  Barcode Cells | **A |**  After reagent loading, priming | Bubbles in any of the reagents?  o **Yes** o **No** | [Top view of DNA cartridge w/o gasket applied] |
| **B |**  After reagent loading, barcoding | Bubbles in any of the reagents?  o **Yes** o **No** | [Top view of DNA cartridge w/o gasket applied] |
| **C |**  After barcoding program finished | Emulsions fully intact and evenly distributed?  o **Yes** o **No** | [8x emulsion-safe tubes w/ barcoded cells] |
| **D |**  After barcoding program finished | Record remaining Barcoding Bead volume [reservoir 7] | [Measure volume] |
| **E |**  After barcoding program finished | Record remaining Barcode Mix volume [reservoir 8] | [Measure volume] |
| **F |**  When removing oil | Total volume ~100 µL in each tube?  o **Yes** o **No** | [8x emulsion-safe tubes w/ barcoded cells and oil removed] |
| **5 |**  Targeted PCR Amplification | **A |**  Before thermal cycling | Thermal cycling protocol correct?  o **Yes** o **No** | [Thermal cycling protocol without tubes, started with remaining time visible] |
| **B |**  Before thermal cycling | PCR skirt used?  o **Yes** o **No** | [Thermal heat block image] |
| **C |**  After thermal cycling | Emulsions fully intact?  o **Yes** o **No** | [8x emulsion-safe tubes w/ barcoded cells] |
| **6 |**  Emulsion Breakage | **A |**  After emulsion breakage | Emulsions fully broken?  o **Yes** o **No** | [8x emulsion-safe tubes w/ broken emulsions] |
| **7 |**  Cleanup PCR Products | **A |**  After enzymatic cleanup | Sample spun down and transferred to new tube after enzymatic cleanup?  o **Yes** o **No** | [Sample after transfer to new tube] |
| **8 |**  PCR Target Library | **A |**  After quantification w/ Qubit (Final Libraries) | DNA concentration within spec?  DNA: > 2.0 ng/µL  Protein: > 2.0 ng/µL  o **Yes** o **No** | [Qubit reading] |
| **B | (Optional)**  Quantification w/ Qubit (Targeted PCR - DNA Library) | DNA concentration recorded?  o **Yes** o **No** | [Qubit reading] |
| **C |**  Bioanalyzer or TapeStation Results | Expected peak at ~460 bp (DNA library) and ~250 (protein library) with few (<5%) off-target fragments (e.g., primer dimers)?  o **Yes** o **No** | [Fragment analyzer results] |